Identification and Evaluation of Sources of Resistance to Stem Rust Race Ug99 in Wheat

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ABSTRACT

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The race Ug99 of *Puccinia graminis* f. sp. *tritici* causing stem rust disease of wheat was initially identified in Uganda in 1998. It was designated as TTKSK based on the North American nomenclature and has caused periodic losses to wheat crops in East Africa. Ug99 has recently moved out of Africa to Yemen and West Asia. The most effective approach to prevent losses from stem rust is through the deployment of resistant cultivars. More effective sources of resistance need to be identified and incorporated in the existing commercial cultivars. The first Stem Rust Resistance Screening Nursery (1stSRRSN) assembled by the International Maize and Wheat Improvement Center (CIMMYT) consisted of elite and advanced CIMMYT bread wheat lines and was evaluated for resistance to Ug99 in Njoro, Kenya for four consecutive seasons (2005 to 2007). Seedling reactions were determined in the greenhouse at the Cereal Disease Laboratory, St. Paul, MN. Two race-specific genes, *Sr24* and *Sr25*, were found to confer resistance to Ug99, although *Sr24* became ineffective to a mutant race of Ug99, TTKST, in 2007. Three previously uncharacterized genes, one each from synthetic wheat, Chinese germplasm, and other genetic backgrounds, were detected. Although 30% of the screened lines were susceptible in the seedling stage, these lines displayed various levels of adult plant resistance (APR) in the field tests. Presence of the APR gene *Sr2*, identified based on the pseudo-black chaff phenotype on glumes and darkened internode, was common in wheat lines with APR. The information on the resistance identified in the 1stSRRSN constitutes an important source for breeding wheat for durable resistance.

Stem or black rust, caused by *Puccinia graminis* Pers. f. sp. *tritici*, has historically caused severe losses to wheat (*Triticum aestivum* L.) production worldwide. Its control for over 30 years through the use of genetic resistance is a remarkable success story. However, in 1998, high susceptibility of International Maize and Wheat Improvement Center (CIMMYT) germplasm was noted in Uganda (9), and an increase in stem rust incidence and severity was observed in Kenya in 2003.

The causal race, commonly known as Ug99 and designated as TTKSK based on the North American nomenclature (2,19), carries virulence for several stem rust resistance genes commonly present in wheat germplasm. Characterization of virulence in the *P. graminis* f. sp. *tritici* populations and germplasm screening were simultane-

ously initiated in Kenya and Ethiopia in 2005. All isolates collected from Kenyan fields in 2004 and 2005 fitted the designation of TTKSK (2,3).

Resistance genes that conferred low reactions to race TTKSK in seedling tests and in the field nursery at Njoro include *Sr13*, -22, -24, -25, -26, -27, -28, -32, -33, -35, -36, -37, -39, -40, -44, and *Tmp* (4). Among the effective genes, *Sr22*, -24, -25, -26, -36, and *Tmp* may have some immediate value (15). The other genes are either located in large alien chromosome segments, requiring a size reduction of the alien segment, or have lacked durability (15). CIMMYT triticale lines, which often carry *Sr27* or *SrSatu*, were highly resistant in the field at Njoro. Although *Sr36* confers resistance to Ug99, another race in the region caused an epidemic on cv. Enkoy, which carries *Sr36*, in 1994 and subsequent years (15). Furthermore, it was found that both *Sr24* and *Sr36* which had shown resistance to Ug99 are no longer effective to new variants of Ug99 (2,5), although they are still very resistant to other important races in other parts of the world. Certain hexaploid synthetic (*T. turgidum* × *Aegilops tauschii*) wheat-

derived advanced lines and some lines with certain Chinese cultivars such as Shanghai no. 7 as parents have also shown adequate levels of resistance. However, the genetic basis of resistance is not known (15,17).

Resistance gene *Sr2* derived from cv. Hope and additional, unknown minor genes, commonly known as the “*Sr2* complex” (7,10,13,14), provided the foundation for durable resistance to stem rust in germplasm from spring wheat of the Northern Great Plains in the United States, Prairie Provinces of Canada, Sydney University in Australia, and the spring wheat germplasm developed by Dr. N. E. Borlaug as part of a program sponsored by the Mexican Government and the Rockefeller Foundation (7,10). Cv. Yaqui 50, released in Mexico during the 1950s, and other *Sr2*-carrying wheat lines released since then, had stabilized the stem rust situation in Mexico and many other countries where modern semidwarf wheat cultivars were adopted. Singh et al. (15) reported that changes in stem rust races have not been observed in Mexico for almost 40 years and natural infections are nonexistent. Another *Sr2*-carrying cultivar, Sonalika, released in the mid-1960s in the Indian subcontinent and subsequently grown on millions of hectares, remained resistant to stem rust. When present alone, the *Sr2* gene confers slow rusting that is not adequate under heavy disease pressure but does provide adequate resistance in combination with other genes. As has been done with leaf rust and yellow rust (16), Singh et al. (14) indicated that adequate levels of multigenic resistance to stem rust can be achieved by accumulating approximately four to five minor genes.

Race-specific stem rust resistance genes can obviously be overcome by new pathotypes relatively rapidly. McIntosh (7) describes cereal rusts as “social” diseases because the individual farmer is subject to airborne spores from areas beyond the farm and part of any inoculum produced on one farm is dispersed to others. Therefore, control of rust diseases must be addressed at the community level, preferably through the widespread use of resistant cultivars. Although several alien genes will provide resistance to Ug99 and its variants,

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the long-term strategy should focus on adult plant resistance (APR). One of the approaches to enhance the resistance conferred by the *Sr2* gene is combining it with three or four additional minor genes to once again achieve durable resistance. The objective of our study was to identify various sources of race-specific resistance and APR to race Ug99 of the stem rust pathogen in CIMMYT-derived wheat germplasm.

MATERIALS AND METHODS

The origin of the 1st Stem Rust Resistance Screening Nursery. In 2005, about 2,000 advanced breeding materials and elite wheat germplasm were evaluated for resistance to race Ug99 of the wheat stem rust pathogen at the Wheat Research Station of Kenyan Agricultural Research Institute, Njoro, Kenya. Based on disease severity and infection responses of the screened lines, the 1st Stem Rust Resistance Screening Nursery (1stSRRSN) was assembled (17). This nursery comprised 103 wheat lines that scored less than 20MS (20% disease severity with a moderately susceptible response) in the field whereas, at the same time, the susceptible genotypes and checks scored between 80S and 100S (80 and 100% disease severity with a susceptible response).

Evaluation of seedling infection types. The 103 lines of the 1stSRRSN were evaluated in the greenhouse at St. Paul, MN using isolate 04KEN156 of *P. graminis* f. sp. *tritici* collected in Kenya in 2004. This isolate was identified as race TTKSK (2,19) based on the 20 differentials in the *P. graminis* f. sp. *tritici* differential set of North America (2,11,12). The procedure of inoculating seedling plants and disease assessment were as described previously (4). Infection types (ITs), described by Stakman et al. (18), were assessed 14 days post inoculation. ITs "0", "1", "2", or combinations thereof were considered low ITs, indicating that the corresponding resistance gene is effective. ITs "3" to "4" were considered high ITs, indicating that the corresponding resistance gene is not effective against the race tested. In each test, 6 to 10 seedling plants were evaluated and the seedling test was repeated once.

The 103 entries of 1stSRRSN were also characterized in the seedling growth stage in the greenhouse of CIMMYT, El Batan, Mexico for their resistance to leaf rust with Mexican *P. triticulturae* races MJB/SP and MFB/SP to postulate the presence of genes *Lr19* and *Lr24* known to be linked to stem rust resistance genes *Sr25* and *Sr24*, respectively. Race MJB/SP is avirulent on genes *Lr19* and *Lr24*, whereas MFB/SP is avirulent on *Lr19* but virulent to *Lr24*. Ten-day-old seedlings, planted as hills of six to eight seeds, were sprayed with urediniospores suspended in Soltrol 170, incubated in a dew chamber for 16 h, and then

placed in a greenhouse maintained at 20 to 23°C. ITs were recorded about 10 days later using a 0-to-4 scale for leaf rust similar to that described by Stakman et al. (18).

Application of molecular markers to detect the presence of resistance genes *Sr24* and *Sr25*. All wheat lines were subjected to molecular marker analysis for the presence of *Sr24* and *Sr25* resistance genes using the co-segregating markers *Sr24* no. 12 and *STSLr19-130*, respectively (14).

Field evaluations for stem rust resistance in Njoro, Kenya. The entries of the 1stSRRSN were tested in 2005, 2006, and two seasons in 2007 as part of a larger field nursery for stem rust screening in Njoro, Kenya established by the Kenyan Agricultural Research Institute in conjunction with the CIMMYT and the Borlaug Global Rust Initiative.

The nursery site is located at 0°20'S, 35°56'E at 2,185 m in elevation, with an average daily minimum temperature of 9.7°C (night) and an average daily maximum temperature of 23.5°C (noon). Variations of average daily temperatures were approximately $\pm 2^\circ\text{C}$, occurring mostly during the day hours when evaluations were carried out. Dew was formed nearly daily.

Entries were planted as two 1-m-row plots during 5 to 15 June in all three main seasons and on 20 December in the 2007 off-season. To facilitate inoculum build-up and uniform dissemination within the nursery, a continuous stem rust spreader row (a mixture of susceptible cvs. Choji and Duma, both carrying *Sr31*) was planted perpendicular to all entries. The spreader rows were inoculated once by dusting them with a mixture of urediniospores and talc powder. The source of inoculum was a bulk of urediniospores collected from experimental plots of Duma in Kenya.

Plant response to rust infection at the adult plant stage was termed "infection response." Based primarily on the size of pustules and the associated necrosis or chlorosis, infection responses were classified into four discrete categories: R = resistant, MR = moderately resistant, MS = moderately susceptible, and S = susceptible (13). Infection responses overlapping between any particular two categories were denoted using a dash. For instance, "MR-MS" denoted an infection response class that overlapped the MR and MS categories, and was sometimes recorded as M by some researchers. Stem rust severity was assessed following the modified Cobb scale (8). Entries were evaluated for infection responses and stem rust severity two to three times between heading and plant physiological maturity. The infection responses and stem rust severity at the soft-dough stage of plant growth (Zadoks scale 8) were used to represent the final disease scores in this report. Due to the close linkage between *Sr2* and pseudo-black chaff

(PBC), scores on PBC were taken to postulate the presence of *Sr2*. This was a visual measure scored as present (+) or absent (-). The data was taken at late milk stage in all the four seasons.

RESULTS

Resistance identified based on seedling ITs. Of the 103 lines tested for seedling IT to Ug99, 71 were resistant, with ITs ranging from 0; to 2+. The genes underlying the resistance were identified as *Sr24*, *Sr25*, and other unknown genes based on the stem rust IT data as well as postulations based on reactions to races MJB/SP and MFB/SP of *P. triticulturae*, the pedigree, and the presence of co-segregating molecular markers. Because genes *Sr24* and *Sr25* are located on alien translocations and inherited together with the leaf rust resistance genes *Lr24* and *Lr19*, respectively, they can be detected easily through tests with leaf rust isolates, information on the pedigree, and molecular marker analysis. The leaf rust tests in Mexico and stem rust tests in St Paul, MN using the race TTKSK were in agreement with the presence of co-segregating molecular markers, and the two genes could be postulated without difficulty. Of the 103 lines, 41 had *Sr24* resistance gene while 18 had *Sr25* (Tables 1 and 2). Resistance in 32 lines was due to APR.

The *Sr24* carrying wheat lines displayed low ITs ranging between ;2= and 12 with *P. graminis* f. sp. *tritici* race TTKSK (Table 1). With *Lr24*-avirulent *P. triticulturae* race MJB/SP, all these lines displayed ITs 0; or ;, similar to the Thatcher tester carrying this gene. However, except for entry no. 6103, all other lines displayed higher ITs ranging between ;1 and 3+ with *Lr24*-virulent race MFB/SP, indicating the presence of *Lr24* and, hence, *Sr24* in these lines. Entry no. 6103 could also carry gene *Lr9*, which confers IT 0; with the two races used. All lines listed in Table 1 were positive for the molecular marker *Sr24* no. 12 that is diagnostic for *Sr24*.

The *Sr25*-carrying wheat lines displayed low ITs ranging between 0; and 1+ with TTKSK and 0; with the two *P. triticulturae* races (Table 2). These lines were positive for the molecular marker *STSLr19-130* and, therefore, postulated to carry the *Sr25* gene.

The remaining 12 seedling resistant lines (Table 3) had an unknown genetic basis because they lacked the molecular markers linked to *Sr24* and *Sr25*, and 11 of these entries did not display low seedling leaf rust ITs. Entry no. 6070 that displayed IT 0; with both *P. triticulturae* races could carry gene *Lr9*. Based on the pedigree information, the unknown resistance genes can be postulated to be derived either from Chinese or synthetic wheat backgrounds (Table 3). Two of the lines had resistance based on sources that remain to be identified. Most of the race-specific resistance

genes, including *Sr24* and *Sr25*, were introgressed from alien sources. The unknown genes derived from crosses with *A. tauschii* could be highly valuable new genetic resources for improvement of resistance against race Ug99.

Thirty-two of the tested lines were susceptible (ITs 3 and 4) at the seedling stage. Such a result is expected for minor genes based on APR (14) and these lines showed from moderate to high levels of resistance at the adult plant stage (Table 4). Twenty-nine of these lines had a PBC phenotype, a trait linked to the *Sr2* gene (Table 4). Absence of PBC expression does not exclude

the possibility that *Sr2* is present in the remaining three wheat lines because, in some genetic backgrounds, PBC remains unexpressed (15).

Resistance identified based on field evaluations. The disease severity was very similar for all the sources of resistance in the first three seasons in Njoro, Kenya (Fig. 1). In the second season of 2007, the disease severity of lines with *Sr24* varied from 15 to 40% (Fig. 1). This was attributed to the appearance in 2006 of a new mutant race (TTKST) that is virulent for gene *Sr24* but remained in a low frequency. This was confirmed by Jin et al.

(2) from samples collected in 2006 and 2007. Seedling tests indicated that the TTKST variant has a virulence pattern identical to race TTKSK, except that it produced high ITs (IT 2+ to 3) on wheat genotypes that carried *Sr24* (2).

Lines with resistance gene *Sr24* showed a higher level of resistance when this gene occurred together with other sources of resistance like the 1A.1R chromosomal translocation from rye. Translocation 1A.1R carries a stem rust resistance gene of unknown relationship to the rye *Sr31* resistance gene and is present in some U.S. winter wheat cultivars such as Amigo,

Table 1. Seedling infection types to *Puccinia graminis* f. sp. *tritici* race Ug99 (TTKSK) and *P. tritricina* races MJB/SP and MFB/SP, field infection responses to race Ug99 for 2005 to 2007 of wheat lines that carry resistance gene *Sr24*, and presence (+) or absence (–) of pseudo-black chaff (PBC) phenotype

Entry no.	Name or cross	Seedling infection type ^a			Year and field rust response ^b					PBC
		<i>P. triticeina</i>		<i>P. graminis</i>	2005	2006	2007-O	2007-M		
		MBJ/SP	MFB/SP	TTKSK						
6001	Krichauff/2*Pastor	;	23c	2-	30 R	5 RMR	5 MSS	40 MSS	+	
6002	Krichauff/2*Pastor	;	3+	2-	20 R	15 RMR	5 MSS	30 MSS	+	
6003	Krichauff/2*Pastor	;	3+	2-	20 R	10 RMR	10 MSS	10 MSS	+	
6004	Krichauff/2*Pastor	;	3	2-	20 R	10 RMR	15 MSS	30 MSS	–	
6005	Krichauff/Pastor		1+3c	2-	5 R	15 M	10 MSS	40 MSS	–	
6006	RL6043/4*Nac//2*Pastor	;	1+	2=	30 R	20 RMR	10 MSS	15 MSS	+	
6007	Sunco/2*Pastor	0;	3+	2-	15 R	15 RMR	10 MSS	10 MSS	+	
6008	Sunco/2*Pastor	;	3+	2-	10 R	30 M	15 MSS	40 MSS	+	
6009	Sunco/2*Pastor	;	3+	2-	10 R	30 M	20 S	50 S	+	
6010	Worrakatta/2*Pastor	;	23c	2-	30 RMR	30 M	20 S	40 MSS	+	
6011	Worrakatta/2*Pastor	;	23c	2-	20 R	20 M	20 S	30 MSS	+	
6012	Worrakatta/2*Pastor	0;	23c	2-	20 R	20 M	20 S	50 MSS	+	
6013	Worrakatta/Pastor	0;	23c	2	30 R	20 M	10 MSS	50 MSS	+	
6014	Worrakatta/Pastor	;	1+3c	2	5 R	20 RMR	5 MSS	40 MSS	+	
6031	TAM200/Pastor//Tobarito97	;	3+	2-	1 R	10 R	10 MSS	6 OS	–	
6073	Ald/CEP75630//CEP75234/PT7219/3/Buc/Bjy/4/Cbrd/5/Tnmu/PF85487	0;	1	2-	10 RMR	10 R	10 MSS	60 M	–	
6074	Ald/CEP75630//CEP75234/PT7219/3/Buc/Bjy/4/Cbrd/5/Tnmu/PF85487	0;	1	2-	5 MR	10 R	10 MSS	60 M	–	
6075	Ald/CEP75630//CEP75234/PT7219/3/Buc/Bjy/4/Cbrd/5/Tnmu/PF85487	0;	1	2-	5 RMR	5 R	10 MSS	50 M	–	
6076	Babax/LR42//Babax	0;	3	2	5MR	30 M	40 S	90 S	–	
6077	Babax/LR42//Babax	;	3+	2-	5 R	10 MR	20 MSS	60 S	+	
6078	Babax/LR42//Babax/3/Babax/LR42//Babax	;	3c3	2-	5 R	10 R	5 RMR	40 M	+	
6079	Bow/Gen//Dern/3/Tnmu	0;	1	2-	5 R	10 RMR	10 MSS	60 S	–	
6081	Bow/Gen//Dern/3/Tnmu	;	1	2-	5 R	10 RMR	5 MSS	50 S	–	
6082	Bow/Gen//Dern/3/Tnmu	;	1	2-	5 RMR	10 R	5 MSS	60 S	–	
6083	Bow/Gen//Dern/3/Tnmu	;	1	2-	5 R	10 RMR	5 MSS	40 MS	–	
6084	Bow/Gen//Dern/3/Tnmu	;	1	2-	5 R	10 M	5 MSS	40 S	–	
6085	CS/Th.cu//Glen/3/Ald/Pvn/4/CS/Le.ra//2*C S/3/CNO79/5/Sirn//Altar84/Ae.sq. (205)/3/3*Buc/6/Pastor	0;	3+	2-	5 R	10 M	20 S	20 MSS	+	
6087	OR 1	0;	3	2-	5 RMR	10 M	20 S	40 S	–	
6088	ORL91256	0;	X	2-	0	10 M	10 MSS	30 MSS	–	
6089	ORL9127	;	3	12	5 RMR	10 M	20 S	40 S	–	
6090	ORL9127/Pastor//Cbrd	;	3	2-	10 RMR	20 M	10 S	40 MS	+	
6091	PBW343/Ducula//PBW343	0;	12	2=	1 R	15 M	10 S	40 S	+	
6092	Thb/Kea//PF85487/3/Milan	;	3	2=	10 RMR	10 M	10 M	10 MS	–	
6093	Thb/Kea//PF85487/3/Milan	;	3	2=	10 RMR	10 M	10 MSS	15 M	–	
6094	Thb/Kea//PF85487/3/Rivadeneira4	;	1	2-	20 S	30 M	20 S	70 S	+	
6095	Tnmu/3/Ald/Coc//Ures	0;	1	2-	15 R	10 M	15 S	40 S	–	
6098	TAM200/Tui//SW89-5124*2/Fasan	;	3	2-	5 R	10 M	10 M	30 M	+	
6099	TAM200/Tui	0;	1	2-	15 MR	10 RMR	20 RMR	30 M	+	
6101	TAM200/Tui	0;	1+	2-	15 MR	10 R	10 RMR	30 M	+	
6102	TAM200/Tui/3/Babax/LR42//Babax	0;	3c3	2-	1 R	15 RMR	5 R	20 MS	+	
6103	EMB16/Cbrd//Cbrd	;	;	2-	5 R	10 RMR	5 R	30 RMR	–	
Check	PBW343	1+3c	3	4	100 S	90 S	100 S	100 S	–	

^a Seedling infection type (IT) for reactions to TTKSK, follow the 0-to-4 scale described in Stakman et al. (18), where ITs 0, 1, 2, and combinations are resistant reactions and ITs 3 and 4 are susceptible reactions.

^b Year 2007-O and 2007-M = off-season and main season, respectively. Field response is a combination of disease severity (0 to 100% of the stem rust infection) using the modified Cobb Scale described in Peterson et al. (8) and infection response follow the description of Roelfs et al. (13), where R = resistant, MR = moderately resistant, M = moderately resistant to moderately susceptible, MS = moderately susceptible, and S = susceptible.

TAM107, and TAM200 (3). This translocation is also present in a CIMMYT spring wheat line TAM200/TUI (14). On average, a reduction of disease severity was also

noted in lines with a combination of *Sr24* and *Sr2* (Table 2). For example, line Krichauff/2*Pastor showed PBC, a phenotypic marker for *Sr2*, and scored 10MSS,

while the same sib-line without PBC scored 40S. Earlier studies had shown that *Sr2* alone will not confer adequate level of resistance to Ug99 (15). Variation in adult

Table 2. Seedling infection types to *Puccinia graminis* f. sp. *tritici* race Ug99 (TTKSK) and *P. tritricina* races MBI/SP and MFB/SP, field infection responses to race Ug99 for 2005-2007 of wheat lines that carry resistance gene *Sr25*, and presence (+) or absence (–) of pseudo-black chaff (PBC) phenotype

Entry no.	Name or cross	Seedling infection type ^a			Year and field rust response ^b				PBC
		<i>P. tritricina</i>		<i>P. graminis</i>	2005	2006	2007-O	2007-M	
		MBJ/SP	MFB/SP	TTKSK					
6015	C80.1/3*Batavia//2*Weebill1	0;	0;	2	10 MR	5 RMR	5 R	10 RMR	+
6016	CMH-81/Hahn//CMH-81/3/Vee/Pjn//2*Tui	0;	0;	2-	5 MR	5 RMR	15 M	10 RMR	+
6017	CMH-81/Hahn//CMH-81/3/Vee/Pjn//2*Tui	0;	0;	;2=	30 R	10 MR	5R	10 RMR	+
6018	CMH83.2517/Pastor	0;	0;	2-	30 RMR	20 M	5 RMR	15 RMR	—
6019	ID#840335//PIN39/Pew/3/Oasis/4*Bor195/4/ Oasis/5*Bor195	0;	0;	2-	10 RMR	5 MR	5 RMR	5 R	+
6021	Kauz*2/Bow//Kauz/3/W98.6.38	0;	0;	2-	10 M	10 RMR	5 R	5 R	+
6022	Oasis/SKauz//4*Bcn/3/2*Pastor	0;	0;	2-	15 RMR	10 RMR	5 R	10 RMR	—
6023	Seri*3//RL6010/4*Yr/3/Pastor	0;	0;	1+	10 M	10 M	5 R	5 RMR	+
6024	Seri*3//RL6010/4*Yr/3/Pastor/4/Bav92	0;	0;	2	10 M	10 M	5 RMR	15 RMR	+
6025	Seri*3//RL6010/4*Yr/3/Pastor/4/Bav92	0;	0;	2-	10 M	10 RMR	5 RMR	10 RMR	+
6026	Seri*3//RL6010/4*Yr/3/Pastor/4/Pastor	0;	0;	2	30 MRMS	20 RMR	5 RMR	10 RMR	+
6027	SKauz/Bav92	0;	0;	2-	10 R	10 MR	1 R	5 RMR	+
6028	SSeri1/Milan	0;	0;	2-	5 RMR	5 R	5 R	5 R	+
6029	Tukuru/Pastor	0;	0;	2-	5 R	5 R	1R	5 R	+
6030	Wheatear	0;	0;	2-	10 RMR	5 R	1 R	5 R	+
6069	Zhengzhou872//Wizza23/Cona-D/3/Sunsu	0;	0;	2-	10 M	10 RMR	5 M	30 M	—
6096	Buc/Pr//Weaver/3/Tukuru	0;	0;	0;	15 RMR	5 M	5 R	10 M	—
6097	CMH-81/Hahn//CMH-81/3/Vee/Pjn//2*Tui	0;	0;	2-	5 MR	5 RMR	5 R	10 MR	+
Check	PBW343	1+3C	3	4	100 S	90 S	100 S	100 S	—

^a Seedling infection type (IT) for reactions to TTKSK, follow the 0-to-4 scale described in Stakman et al. (18), where ITs 0, 1, 2, and combinations are resistant reactions and ITs 3 and 4 are susceptible reactions.

^b Year 2007-O and 2007-M = off-season and main season, respectively. Field response is a combination of disease severity (0 to 100% of the stem rust infection) using the modified Cobb Scale described in Peterson et al. (8) and infection response follow the description of Roelfs et al. (13), where R = resistant, MR = moderately resistant, M = moderately resistant to moderately susceptible, MS = moderately susceptible, and S = susceptible.

Table 3. Seedling infection types to *Puccinia graminis* f. sp. *tritici* race Ug99 (TTKSK) and *P. tritricina* races MBI/SP and MFB/SP, field infection responses to race Ug99 for 2005-2007 of wheat lines that carry unknown resistance genes, and presence (+) or absence (–) of pseudo-black chaff (PBC) phenotype

Entry no.	Name or cross	Seedling infection type ^a			Year and field rust response ^b				PBC	Source ^c
		<i>P. tritricina</i>		<i>P. graminis</i>	2005	2006	2007-O	2007-M		
		MBJ/SP	MFB/SP	TTKSK						
6064	Chil/Chuan Mai 18/4/Buc/Bjy/3/Cndr/Ana//Cndr/Mus	23c	;1	1+3C	30 M	30 MSS	20 MSS	30 M	...	Chuan Mai 18
6065	Kauz//Altar84/Oasis/3/Kauz/4/SW94.15464	3c	;123	2++	20 RMR	15 M	10 R	30 RMR	+	Durum/Chinese
6066	Milan/Shanghai 7/3/Thb/CEP7780//Shanghai 4/Lira/4/Shanghai 4/Chil	3c	;1	2+	20 MRMS	5 R	5 R	10 RMR	—	Chinese
6067	Ning Mai 50	;12	;	2++	30 RMR	40 M	20 MR	40 MR	—	Ning Mai 50
6068	Ning Mai 9415.16//Shanghai 4/Chilero/3/Ning Mai 50	23c	;	2+	30 RMR	10 M	5 RMR	10 RMR	—	Chinese
6070	Chen/Ae.sq.//Bcn/3/CMH81.38/2*Kauz	0;	0;	1+3C	5 R	10 MR	40 MR	60 MSS	—	Synthetic
6071	Chen/Ae.sq.//2*Weaver/3/Oasis/5*Bor195	3+	3	2+3C	10 MSS	30 M	5 RMR	10 MR	+	Synthetic
6072	Chibia/Dulus	1+3c	;1-	2+	30 MRMS	10 M	5 RMR	30 MR	—	Dulus
6105	Ning Mai 9558//Chilero/Chuan Mai 18	1+3c	;	2-	5 R	5 RMR	10 RMR	10 RMR	—	Chinese
6106	Yang87-158*2//Milan/Shanghai 7	3	;	2	30 RMR	5 RMR	15 RMR	20 RMR	+	Shanghai 7
6107	Croc_1/Ae..sq.(205)//Fct/3/Pastor	3+	3	2-	10 R	10 R	10 R	5 R	+	Synthetic
6108	Croc_1/Ae.sq.(205)//Fct/3/Pastor	;12	;	2-	15 M	20 M	5 R, 40 S	5 RMR	+	Synthetic
Check	PBW343	1+3C	3	4	100 S	90 S	100 S	100 S	—	...

^a Seedling infection type (IT) for reactions to TTKSK, follow the 0-to-4 scale described in Stakman et al. (18), where ITs 0, 1, 2, and combinations are resistant reactions and ITs 3 and 4 are susceptible reactions.

^b Year 2007-O and 2007-M = off-season and main season, respectively. Field response is a combination of disease severity (0 to 100% of the stem rust infection) using the modified Cobb Scale described in Peterson et al. (8) and infection response follow the description of Roelfs et al. (13), where R = resistant, MR = moderately resistant, M = moderately resistant to moderately susceptible, MS = moderately susceptible, and S = susceptible.

^c Probable source parent.

plant stem rust severity of lines that carried *Sr2* supports this observation.

Genes *Sr24* and *Lr24*, conferring resistance to stem and leaf rust, respectively, are located on chromosome 3DL on a translocated segment originating from *Thinopyron elongatum*. There are three distinct *Sr24*-carrying translocations: the original translocation carried a gene for red grain color, the shorter segment with white grain, and a third segment where a very small segment has been retranslocated onto chromosome 1BS (14). In all three segments, both *Sr24* and *Lr24* are present together. Therefore, selection for *Lr24* with an avirulent leaf rust isolate can be used as an indirect selection strategy for *Sr24*. The *Sr24* resistance gene had earlier looked like an attractive candidate for future breeding efforts; however, virulence to *Sr24* is known in South Africa and India (14) and the recent detection of the Ug99 mutant race TTKST which is virulent on *Sr24* reduces its usefulness in breeding (2).

Sources with the *Sr25* gene showed low disease severity across the four seasons and, in fact, very low severity in the 2007 off-season. It was also noted that *Sr25*

conferred a high level of resistance only in some genetic backgrounds, especially when the slow rusting APR gene *Sr2* was also present (e.g., in lines 'Super Seri no. 1' [which has yellow flour] and 'Wheatear' [which has white flour]). However, a virulent race was detected in India in 2008 (1) but CIMMYT wheat lines with *Sr25* in the 1stSRRSN remain resistant due to the presence of other unknown resistance genes. The *Sr25* gene is located on a *T. elongatum* translocation together with leaf rust resistance gene *Lr19* on chromosome 7DL (14). A white-floured mutant of the translocation, developed by D. R. Knott, was transferred into some Australian and CIMMYT wheat backgrounds. Almost all the tested lines with *Sr25* of CIMMYT origin also carried PBC and, hence, *Sr2* (Table 1). Therefore, the combination of *Sr25* with *Sr2* may be the reason for low disease severity in these entries. However, due to the major gene effect of *Sr25*, it will be difficult to quantify how much resistance *Sr2* could be contributing. Thus, the *Sr25* gene could be a good resistance resource but should be used in combination with other effective race-

specific resistance genes as well as *Sr2* or other minor genes.

The unknown resistance sources included wheat lines that were derived from crosses with synthetic wheat lines or Chinese wheat genotypes (Table 3). Two of the lines had unknown and not-easy-to-predict sources of resistance. The lines with the Chinese parents "Milan/Sh4/3/Thb/CEP7780/Sh4/Lira/4/Sh4/Chil", "Ning Mai 9415.16/Sh4/Chil/3/Ning Mai 50", and "Ning Mai 9558/Chil/Chum18" scored, on average, 10% severity with an R-MR type of infection response. The gene responsible for the resistance is likely derived from Shanghai no. 7 or Shanghai no. 4 and is tentatively called *SrSha7*. The CIMMYT parents in the crosses with the Chinese resistant sources are likely to be susceptible to Ug99. Further studies are needed to track the source of resistance in the Chinese wheat parents.

The resistance observed in the synthetic genetic background was due to *A. tauschii* or *Triticum durum* parents used in the development of synthetic wheat lines (15). The stem rust severity was between 5 and 10%, with infection responses of R and

Table 4. Seedling infection types and field infection responses for 2005 to 2007 for wheat lines that carry adult plant resistance genes to race Ug99 (TTKSK) of *Puccinia graminis* f. sp. *tritici* and presence (+) or absence (–) of pseudo-black chaff (PBC)

Entry no.	Name or cross	IT ^b	Year and field rust response ^a				PBC
			2005	2006	2007-O	2007-M	
6032	Babax/3/Oasis/SKauz/4*Bcn/4/Pastor	4	30 MSS	30 MSS	40 S	10 MSS	+
6033	Cndo/R143/Ente/Mexi_2/3/Ae.sq./4/Weaver/5/2*Kauz/6/Fret2	4	30 MRMS	20 M	40 MSS	30 MSS	+
6034	Cndo/R143/Ente/Mexi_2/3/Ae.sq./4/Weaver/5/2*Kauz/6/Fret2	4	30 MRMS	20 M	20 MSS	20 MSS	+
6035	Dollarbird	3	10 R	15 M	15 M	20 M	+
6036	Fret2*2/Kukuna	4	30 MS	20 M	30 MSS	30 M	+
6037	Hartog	3+	10 MR	10 M	10 M	10 M	+
6038	He1/3*Cno79/2*Seri/3/Attila/4/WH542	3	10 R	50 MSS	20 MSS	40 MSS	–
6039	Hpo/Tan/Vee/3/2*Pgo/4/Milan/5/SSeri1	3+	30 MR	40 MSS	30 MSS	10 MSS	+
6041	Kamb1*2/Khvaki	3+	30 MS	20 M	15 M	10 MSS	+
6042	Kiritati	3+	10 R	15 M	15 M	10 MS	+
6043	Parus	4	10 R	40 S	15 M	20 MSS	+
6044	Pastor/Milan	3+	30 M	15 S	15 S	5 S	–
6045	Pavon F 76	3+	10 R	15 M	10 M	5 M	+
6046	Pfau/Weaver*2/Kiritati	3+	20 MRMS	20 M	10 MS	5 MS	+
6047	Pfau/Weaver*2/Kiritati	3+	20 MRMS	20 M	10 MS	10 MSS	+
6048	Pfau/Weaver*2/Kiritati	3+	20 MRMS	30 M	10 MSS	5 MSS	+
6049	Pfau/Weaver*2/Kiritati	3+	20 MRMS	15 M	5 MS	5 MSS	+
6050	Pfau/Weaver*2/Kiritati	3+	20 MRMS	10 M	10 MS	10 MSS	+
6051	Pfau/Weaver/Kiritati	3+	20 MRMS	10 M	10 MSS	5 MSS	+
6052	Pfau/Weaver/Kiritati	3	20 MRMS	20 M	10 MSS	10 MSS	+
6053	Pfau/Weaver/Kiritati	3+	15 MRMS	10 M	10 MSS	10 MSS	+
6054	Pfau/Weaver/Kiritati	3+	20 MRMS	20 M	20 S	40 MSS	+
6055	Pgo/Croc_1/Ae.sq.(224)/3/2*BorIL95/4/Circus	3+	10 MRMS	5 M	15 MSS	20 MSS	+
6056	Pgo/Croc_1/Ae.sq.(224)/3/2*BorIL95/4/Circus	4	10 MRMS	10 M	10 MSS	10 MS	+
6057	Pgo/Seri/Bav92	3+	10 R	15 M	10 MSS	10 MSS	+
6058	Pvn/Car422/Ana/5/Bow/Crow/Buc/Pvn/3/Yr/4/Trap#1	3+	10 R	30 M	5 MS	10 MS	+
6059	Kingbird	3+C	5 MRMS	5 MR	5 MSS	5 MSS	+
6061	Kingbird	3+C	5 MRMS	5 M	5 MSS	5 MSS	+
6062	Temporalera M 87*2/4/HD2281/Trap#1/3/Kauz*2/Trap/Kauz	3	30 M	20 M	10 M	30 MSS	–
6063	WL6736/5/2*BR12*3/4/IAS55*4/CI14123/3/IAS55*4/EG,AUS/IAS55*4/Ald/6/Oasis/5*BorIL95/7/BorIL95	3+	20 MSS	20 M	5 MS	10 MSS	+
6086	Esda/Lira/Milan/3/Vee#5/Sara	3	20S	40 S	20 S	20MSS	+
6104	Juchi F2000	4	20 RMR	20 M	15 MSS	30 M	+
Check	PBW343	4	100 S	90 S	100 S	100 S	–

^a Year 2007-O and 2007-M = off-season and main season, respectively. Field response is a combination of disease severity (0 to 100% of the stem rust infection) using the modified Cobb Scale described in Peterson et al. (8) and infection response follow the description of Roelfs et al. (13), where R = resistant, MR = moderately resistant, M = moderately resistant to moderately susceptible, MS = moderately susceptible, and S = susceptible.

^b Seedling infection type (IT) for reactions to TTKSK, follow the 0-to-4 scale described in Stakman et al. (18), where ITs 0, 1, 2, and combinations are resistant reactions and ITs 3 and 4 are susceptible reactions.

MR. There were some exceptions where the field scores were higher, especially in the main season of 2007 (Table 3). Resistance in synthetic-wheat-derived lines can be due to the presence of *Sr13* and *Sr14* on chromosome 6AL and 1BL, respectively, originating from durum wheat parents of synthetic wheat lines and *Sr33* and a recently designated resistance gene *Sr45* on chromosomes 1DL and 2DS, respectively, deriving from *A. tauschii* parents. Genes *Sr13*, *Sr14*, and *Sr33* confer only moderate levels of resistance (4) and they will be useful in areas where stem rust pressure remains at moderate levels. Virulence to both *Sr13* and *Sr14* is known among races different from Ug99 (14). Response of *Sr45* with Ug99 in seedling and field conditions is yet to be determined.

Although the disease severity of lines with APR was, on average, higher than that of the lines with other resistance sources (Fig. 1), it maintained a constant infection level (15 to 20%) throughout the testing period of multiple seasons. Most of the lines in this group scored as low as 5% severity, on average, with variable infection responses. Kingbird, although susceptible as seedlings in greenhouse tests, was the best line for APR across seasons and maintained very low severity during all seasons. It also carried *Sr2* based on the presence of PBC.

It has been found earlier (6) that *Sr2* is closely linked to PBC. Of the 32 lines showing APR, almost 90% had the PBC phenotype, indicating that they carried *Sr2* (Table 4). The slow rusting gene *Sr2* confers only moderate levels of resistance when present alone. On wheat lines in 1stSRRSN that displayed PBC, we observed varying degrees of disease, with a maximum severity reaching up to 40% in some seasons compared with 90% severity for highly susceptible material. Infection responses varying from MR to S were scored on the same internodes of *Sr2*-carrying plants (Table 4). On several other wheat materials (not included in the 1stSRRSN and *data not presented*), we also have observed that the level of resistance conferred by *Sr2* under high disease pressure in Kenya was not sufficient. This indicated that the resistant lines with APR contained other genes, most likely minor, in addition to gene *Sr2*. Genetic analyses of resistance in selected APR wheat cultivars are currently underway to unravel the APR to stem rust.

DISCUSSION

Based on the seedling ITs as well as field disease severity and infection responses, we identified a number of resistance genes that are effective against Ug99. Although *Sr24* had initially shown a high level of resistance to Ug99, it was

later overcome by the TTKST mutant of Ug99. *Sr24* conferred increased resistance in the presence of the slow rusting APR gene *Sr2*. *Sr25* showed resistance across the four seasons to Ug99 and its enhanced expression in CIMMYT germplasm was probably due to the presence of *Sr2* or other minor genes. The three possibly new sources of resistance from synthetic, Chinese, and other unknown origins represent highly valuable genetic resources in achieving resistance to Ug99. Wheat material with high levels of APR should be used for developing wheat cultivars with durable resistance. The contribution of *Sr2* to APR is obviously highly relevant, given that PBC, a trait associated with *Sr2*, was present in most of the APR wheat lines as well as several of the wheat lines that carried race-specific resistance. We propose to genetically dissect the APR found in these lines to better understand and apply it in breeding for durable stem rust resistance.

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Fig. 1. Average field stem rust severity score (%) on sources of resistance over four seasons (2005 to 2007) conducted in Njoro, Kenya.

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